

# ANTIBACTERIAL ACTIVITY OF TITANIUM DIOXIDE AND AG-INCORPORATED DLC THIN FILMS

Veronika Vymětalová<sup>1</sup>, Miroslav Jelínek<sup>1</sup>, Petr Písařík<sup>1</sup>, Jan Mikšovský<sup>1</sup>,  
Jan Remsa<sup>1</sup>, Veronika Řasová<sup>1</sup>

<sup>1</sup>Czech Technical University in Prague, Faculty of Biomedical Engineering,  
Kladno, Czech Republic

## Abstract

Titanium dioxide (TiO<sub>2</sub>) and Ag-incorporated diamond-like carbon (DLC) films were prepared on different substrates. The films were prepared by pulsed laser deposition (PLD). TiO<sub>2</sub> and Ag were selected due to their potential values as biomaterials. Silver is effective against a wide range of spectrum including Gram-negative and Gram-positive bacteria and yeast. TiO<sub>2</sub> and Ag-incorporated DLC thin films are suitable candidates for application on biomedical devices and implants due to their biocompatibility, chemical inertness, and mechanical properties. Thin films are widely used in coronary artery stents, dental implants, heart valves and other vascular devices. The microstructure and antibacterial properties of TiO<sub>2</sub> and silver-doped diamond-like carbon (DLC) films have been investigated. The films structural quality was evaluated using SEM microscopy, AFM microscopy and Raman spectroscopy. The antibacterial activity was determined using Gram-negative bacteria *Escherichia coli* and Gram-positive bacteria *Bacillus subtilis*. Our results demonstrate that the TiO<sub>2</sub>, nitrogen doped titanium oxides TON and Ag-incorporated DLC films are potentially useful as biomedical materials having good antibacterial properties.

## Keywords

thin films, pulsed laser deposition, antibacterial properties, Gram-negative bacteria, Gram-positive bacteria, implants

## Introduction

Titanium dioxide (TiO<sub>2</sub>) is an important material for many industrial applications. The photocatalytic properties of titanium dioxide were discovered by Akira Fujishima in 1967 and published in 1972. When a photocatalyst TiO<sub>2</sub> captures ultraviolet light (UV) either from sun or fluorescent light, it forms activated oxygen from water or oxygen in the air. The process on the surface of TiO<sub>2</sub> was called the *Honda-Fujishima effect*. This process is similar to photosynthesis. The formed activated oxygen is strong enough to oxidize and decompose organic materials (kill bacteria and fungi) and destroy single-celled organism. The very strong oxidizing power of photocatalytic nano particle titanium dioxide can destroy procaryotic bacteria's cell membrane causing leakage of the cytoplasm, which inhibits bacteria's metabolism and ultimately results in the death and decomposition of bacteria. Photocatalytic and antibacterial properties of TiO<sub>2</sub> thin films were often studied [1, 2, 3, 4, 5]. TiO<sub>2</sub> thin films have been used in clinical applications for their ability to kill many microorganisms. Biomedical devices with TiO<sub>2</sub>

coating decreased hospital associated infections, bacterial colonization and proliferation. Photocatalysis has been shown to be capable of killing more Gram-negative than Gram-positive bacteria [6]. In this study, we focused on preparation of nitrogen doped titanium oxides films (TON) and study their antibacterial activity. It has been reported that titanium dioxide, when doped with nitrogen ions, is also a photocatalyst under either visible or UV light [7].

DLC films are suitable biomaterials for many biomedical applications and devices including bone implants and cardiovascular devices. Silver ions have been widely used as an antibacterial agent in biomedical engineering [8]. The antibacterial effect of silver is related to blockage of the microbial DNA. Some authors associate the toxic effect of silver leading to cell death with inactivation of enzymes [9, 10]. Silver is effective against a wide range of spectrum including Gram-negative and Gram-positive bacteria and yeast. We investigated the antibacterial properties Ag incorporated DLC films. Ag doped DLC films has been suggested to be potentially useful in biomedical devices.

## Material and methods

Thin films were prepared by pulsed laser deposition technique (PLD) with a KrF excimer laser Compex Pro 205 F with energy density ranged from  $3.2 \text{ J}\cdot\text{cm}^{-2}$  to  $9 \text{ J}\cdot\text{cm}^{-2}$ ,  $\lambda = 248 \text{ nm}$ , the number of laser pulses varied from 6 000 to 27 500, repetition frequency set to 10 Hz. Deposition pressure varied from 2 Pa to 20 Pa with atmosphere composed of  $\text{O}_2$  or  $\text{N}_2$ . Thin layers were deposited onto silicon wafers Si (111) for  $\text{TiO}_2$  and TON films, and Si (100) for Ag doped DLC or chemical glass, all  $10\times 10 \text{ mm}$ . More detailed information on deposition conditions of thin films can be found in previous studies [11, 12].

The  $\text{TiO}_2$ , TON and DLC antibacterial activity was determined using Gram-negative bacteria *Escherichia coli* and Gram-positive bacteria *Bacillus subtilis*. *B. subtilis* is a Gram-positive bacterium commonly found in soil, while *E. coli* is a Gram-negative bacterium found in large intestine of mammals. The microbial *Escherichia coli* cells were grown in nutrient media LB and *Bacillus subtilis* cells in nutrient media LB and MPB (pepton, beef extract, NaCl, pH 7.0). For the preparation of the solid media, the nutrient media were supplemented with 2% bacteriological agar as a solidifying agent. The tested  $\text{TiO}_2$ , TON and Ag incorporated DLC thin films were placed into sterile chambers and covered with 1 ml of the overnight *E. coli* and *B. subtilis* cultures diluted to a cell concentration of  $10^6 \text{ CFU}\cdot\text{ml}^{-1}$ . The sterile chambers were cultivated at  $35^\circ\text{C}$  and aliquots of  $100 \mu\text{l}$  suspension were taken and diluted to  $10^{-6}$  after 8 h. Afterwards  $100 \mu\text{l}$  of each diluted sample were spread onto agar plates, incubated for 48 h and then the number of the growing colonies was counted.

## Results

Thin films morphology and topology were studied by scanning electron microscopy (SEM) and atomic force microscopy (AFM). The thickness of titanium dioxide layers was between 0.1 and  $10 \mu\text{m}$  depending on the background pressure and number of laser pulses. The photocatalytic properties of  $\text{TiO}_2$  layers were determined by 4-chlorine phenol (4CIP) solution ( $3\times 10^{-3} \text{ mol}$ ) degradation. This method has been adapted [11]. Each sample was put into a Suprasil cell (3.5 ml volume) with magnetic stirring and were continuously illuminated by a mercury flash lamp ( $8.75 \text{ mW}/\text{cm}^2$ ). To prevent the decomposition of molecules by UV irradiation alone, filtering wavelengths shorter than 360 nm were implemented. Additionally, an IR filter was used to prevent the cell heating. The cell temperature was lower than  $30^\circ\text{C}$ . The illuminated area was roughly  $70 \text{ mm}^2$  and the sample plane was set to be perpendicular to the light

source. The pH decrease over time expresses photocatalytic properties of the sample. For the calculation of final pH change, the pH value from the 100<sup>th</sup> minute was taken and subtracted from the value from the 20<sup>th</sup> minute after the irradiation started.

The photocatalytic properties of amorphous, rutile/anatase and anatase phases of  $\text{TiO}_2$  films are summarized in Table 1.

Tab. 1: Crystalline structure and photocatalytic activity

| Sample           | Crystalline structure (XRD) | pH change (%) |
|------------------|-----------------------------|---------------|
| $\text{TiO}_2$ A | Amorphous                   | 2.3           |
| $\text{TiO}_2$ B | Not measured                | 7.36          |
| $\text{TiO}_2$ C | Rutil, Anatase weak         | 8.54          |
| $\text{TiO}_2$ D | Anatase                     | 8.36          |
| $\text{TiO}_2$ E | Anatase, Rutile             | 3.92          |

The bacterial efficacy (BE) of the  $\text{TiO}_2$ , TON and Ag incorporated DLC thin films on *E. coli* and *B. subtilis* strain was estimated according to the formula:

$$\text{BE} = \frac{N_{\text{ref}} - N_{\text{exp}}}{N_{\text{ref}}} \cdot 100,$$

where:

$N_{\text{ref}}$  – alive number in reference group, number of bacteria on the control samples,

$N_{\text{exp}}$  – alive number in experiment group, number of bacteria on the samples.

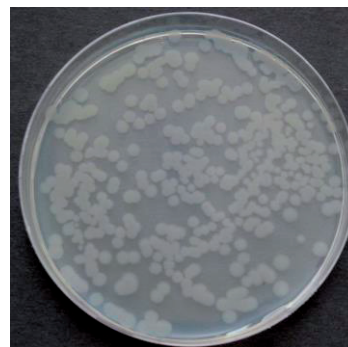
As a control or reference sample a clean substrate was used. The reference group in this case is an amount of bacteria that grew up/perished on this substrate.

Tab. 2: Bacterial efficacy of *B. subtilis*.

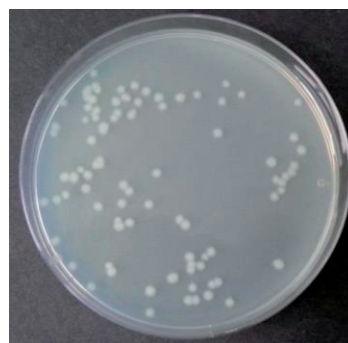
| Sample           | BE (after 1.5 h) | BE (after 3 h) |
|------------------|------------------|----------------|
| $\text{TiO}_2$ A | 34%              | 69%            |
| $\text{TiO}_2$ B | 63%              | 86%            |
| $\text{TiO}_2$ C | 60%              | 83%            |
| $\text{TiO}_2$ D | 69%              | 90%            |
| $\text{TiO}_2$ E | 57%              | 72%            |

Tab. 3: Bacterial efficacy of *B. subtilis*.

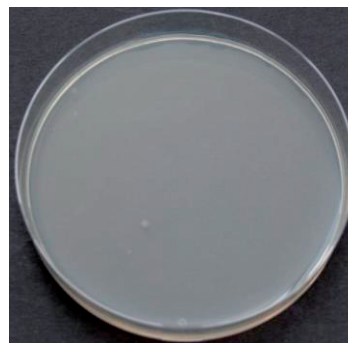
| Sample | BE (after 1.5 h) | BE (after 3 h) |
|--------|------------------|----------------|
| TON 1A | 87%              | 93%            |
| TON 1B | 68%              | 99%            |
| TON 2A | 76%              | 100%           |
| TON 2B | 41%              | 100%           |
| TON 4A | 69%              | 95%            |
| TON 4B | 63%              | 95%            |

Tab. 4: Bacterial efficacy of *E. coli*.

| Sample             | BE (after 1.5 h) | BE (after 3 h) |
|--------------------|------------------|----------------|
| TiO <sub>2</sub> A | 59%              | 92%            |
| TiO <sub>2</sub> B | 84%              | 100%           |
| TiO <sub>2</sub> C | 80%              | 100%           |
| TiO <sub>2</sub> D | 91%              | 100%           |
| TiO <sub>2</sub> E | 61%              | 95%            |

Tab. 5: Bacterial efficacy of *E. coli*.

| Sample | BE (after 1.5 h) | BE (after 3 h) |
|--------|------------------|----------------|
| TON 1A | 89%              | 97%            |
| TON 1B | 79%              | 99%            |
| TON 2A | 85%              | 100%           |
| TON 2B | 92%              | 100%           |
| TON 4A | 92%              | 100%           |
| TON 4B | 89%              | 97%            |

Fig. 1: Antibacterial properties of TON 2 thin film against *Bacillus subtilis*: control sample; after 90 and after 180 minutes.Tab. 6: Bacterial efficacy of *B. subtilis*.

| Sample  | BE (after 1.5 h) | BE (after 3 h) |
|---------|------------------|----------------|
| AgDLC-6 | 88%              | 93%            |
| AgDLC-7 | 85%              | 93%            |
| AgDLC-8 | 96%              | 99%            |

Tab. 7: Bacterial efficacy of *E. coli*.

| Sample  | BE (after 1.5 h) | BE (after 3 h) |
|---------|------------------|----------------|
| AgDLC-6 | 99%              | 99%            |
| AgDLC-7 | 99%              | 98%            |
| AgDLC-8 | 28%              | 98%            |

Antibacterial properties of TiO<sub>2</sub> and TON layers against *Bacillus subtilis* and *Escherichia coli* were measured. The antibacterial effects of TiO<sub>2</sub> and TON thin films against *Bacillus subtilis* are shown in Tab. 2, 3 and Fig. 1, against *Escherichia coli* in Tab. 4 and 5. Antibacterial activity TiO<sub>2</sub> films (*B. subtilis*) was from 34% to 69% after 1.5 h resp. from 69% to 90% after 3 h. Bacterial efficiency TON films (*B. subtilis*) was from 41% to 87% after 1.5 h resp. from 93% to 100% after 3 h. Antibacterial activity TiO<sub>2</sub> films (*E. coli*) was from 59% to 91% after 1.5 h resp. from 92% to 100% after 3 h. Antibacterial activity TON films (*E. coli*) was from 85% to 92% after 1.5 h resp. from 97% to 100% after 3 h. The antibacterial effects of Ag incorporated DLC thin films against *Bacillus subtilis* and *Escherichia coli* are shown in Tab. 6, 7 and Fig. 1.

## Discussion

The main finding of the study is the high photocatalytic and antibacterial activity of rutile and anatase TiO<sub>2</sub> layers. Their photocatalytic properties were tested by the measurement of pH change. Thin films showed excellent antibacterial activity, whereas amorphous TiO<sub>2</sub> layers were also photocatalytically active but with low antibacterial activity. Ag incorporated DLC thin layers showed excellent antibacterial activity in this study. The highest antibacterial efficiency was reached by nitrogen doped TiO<sub>2</sub> thin films. It was found that the Gram-negative bacterium *Escherichia coli* is more sensitive to antibacterial effects of TiO<sub>2</sub> and Ag-incorporated DLC thin films than Gram-positive bacterium *Bacillus subtilis* [6]. The difference between Gram-positive (*B. subtilis*) and Gram-negative (*E. coli*) bacteria is due to bacterial cell wall structure and peptidoglycan layers thickness.

## Conclusion

In this paper, we investigated the antibacterial activity of TiO<sub>2</sub> thin films and nitrogen doped TiO<sub>2</sub> thin films produced by using pulsed laser deposition technique and antibacterial activity of Ag incorporated DLC films prepared by pulsed laser deposition (PLD).

The influence of deposition parameters on physical and biological properties was studied. Our results proved that the highest antibacterial efficiency can be reached by nitrogen doped TiO<sub>2</sub> thin films. In fact all kinds of prepared thin films can be useful in biomedical devices and surgical instruments having good antibacterial characteristics for testing Gram-negative bacteria *Escherichia coli* and Gram-positive bacteria *Bacillus subtilis*.

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Veronika Vymětalová  
Department of Natural Sciences  
Faculty of Biomedical Engineering  
Czech Technical University in Prague  
nám. Sítná 3105, CZ-272 01 Kladno

E-mail: vymetalova@fbmi.cvut.cz  
Phone: +420 224 358 496